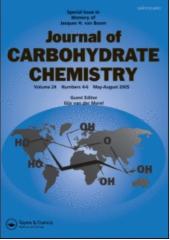
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SYNTHESIS OF A MANNOTETRAOSE—THE REPEATING UNIT OF THE CELL-WALL MANNANS OF *MICROSPORUM GYPSEUM* AND RELATED SPECIES OF *TRYCHOPHYTON*

Linsen Heng^a; Jun Ning^a; Fanzuo Kong^a ^a Research Center for Eco-Environmental Science, Academia Sinica, Beijing, China

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SYNTHESIS OF A MANNOTETRAOSE—THE REPEATING UNIT OF THE CELL-WALL MANNANS OF *MICROSPORUM GYPSEUM* AND RELATED SPECIES OF *TRYCHOPHYTON*

Linsen Heng, Jun Ning, and Fanzuo Kong

Research Center for Eco-Environmental Science, Academia Sinica. P.O. Box 2871, Beijing 100085, China

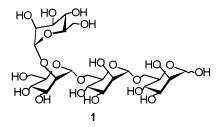
ABSTRACT

A tetrasaccharide, α -D-mannopyranosyl- $(1\rightarrow 2)$ - α -D-mannopyranosyl- $(1\rightarrow 6)$ - α -D-mannopyranosyl- $(1\rightarrow 6)$ -D-mannopyranose (1), the repeating unit of the cell-wall mannans of *Microsporum gypseum* and related species of *Trychophyton*, was synthesized using 6-*O*-acetyl-2,3,4-tri-*O*-benzoyl- α -D-mannopyranosyl trichloroacetimidate (5) and 2-*O*-acetyl-3,4,6-tri-*O*-benzoyl- α -D-mannopyranosyl trichloroacetimidate (13) as the glycosyl donors in "the inverse Schmidt" procedure.

INTRODUCTION

Dermatophytes are a group of fungi which parasitise man and animals, and cause superficial cutaneous infections involving primarily the keratinised tissues of the epidermis, nails, and pilosebaceous follicles. The cell wall of dermatophytes is made up of chitin, a water-insoluble β -(1 \rightarrow 3)-glucan, and water-soluble polysaccharides which are antigenically relevant. In many ascoycetous yeasts,¹ the antigenically relevant outer polysaccharides have a skeleton composed of α -(1 \rightarrow 6)linked mannopyranosyl residues, to most of which are attached one branching moiety, each composed of chains of various lengths containing mainly α -, β (or both)-(1 \rightarrow 2) and/or $-(1\rightarrow3)$ links (the so-called comb-like structure, as for instance, in *Candida maltosa* and *C. Tropicalis*²). A more complex structure (a treelike structure) has been reported for *C. Albicans*,^{3,4} which shows an analogous backbone of α -(1 \rightarrow 6)-linked mannopyranosyl residues with (1 \rightarrow 2) and/or (1 \rightarrow 3) side

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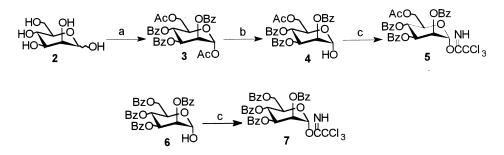


chains, some of which are branched. A third type of sequence has been proposed for the *C*. *Krusei* mannan,⁵ which contains a main chain of mannopyranose with $(1\rightarrow 2)$ and $(1\rightarrow 6)$ linkages in a 3:1 ratio but is lightly branched, either at the 2- or 6-positions.

In 1995, Jiménez-Barbero et al.⁶ investigated the structures of cell-wall mannans isolated from *Microsporum gypseum* and related species of *Trychophyton* and found that all of them consist of an α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 6)- α -D-mannopyranosyl-(1 \rightarrow 6)-D-mannopyranose repeating tetrasaccharide. Syntheses of the fragments of polysaccharides are important for elucidation of biological functions of polysaccharides. In this paper, as a part of our continuous synthetic approach directed toward epitopes of fungal cell-wall mannans, we report the synthesis of a tetrasaccharide 1, the repeating unit of mannans isolated from *Microsporum gypseum* and related species of *Trychophyton*.

RESULTS AND DISCUSSION

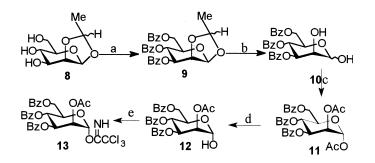
Tritylation of mannose (2) followed by benzoylation in a one-pot manner gave the 1,2,3,4-tetra-O-benzoyl-6-O-trityl-D-mannopyranose, selective acetolysis of which using CH₂Cl₂-AcOH-Ac₂O-H₂SO₄ in a ratio of 1:1:0.6:0.175 afforded the corresponding 1,6-diacetate **3** in 71.3% yield (for three steps). The diacetate **3** was selectively deacetylated at the anomeric position with benzylamine in THF in high yield to give the corresponding 6-O-acetyl-2,3,4-tri-O-benzoyl-D-mannopyranose (**4**). Subsequent reaction of **4** with CCl₃CN/DBU in dichloromethane afforded the glycosyl donor **5**. A similar procedure gave another glycosyl donor **7** (Scheme 1).



Scheme 1. Reagents and conditions: (a) i.trityl chloride (1.2 equiv), pyridine, 50°C, 32 h; ii. Ph-COCl (4.8 equiv), $<40^{\circ}$ C, 24 h; iii.CH₂Cl₂/HOAc/Ac₂O/H₂SO₄ = 1/1/0.6/0.175 (v/v), rt, 20 h, 71.3% (for three steps); (b) benzylamine (3.2 equiv), THF, rt, 24 h, 86.2%.(c) CCl₃CN (2.3–3.2 equiv), DBU (0.18–0.23 equiv), rt, 5 h, 88.1%–85.6%.



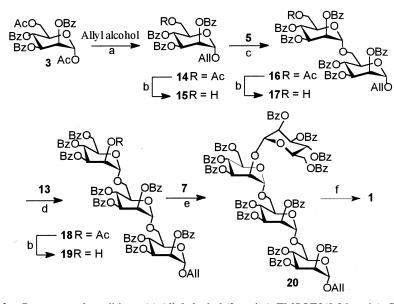
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Scheme 2. Reagents and conditions: (a) PhCOCl (3.3 equiv), pyridine, $<40^{\circ}$ C, 24 h, 94.8%; (b) F₃CCOOH (90%), rt, 4 h, 89.0%; (c) (Ac)₂O, pyridine, rt, 5 h, 100%; (d) benzylamine (4.0 equiv), THF, rt, 24 h, 88.4%.(e) CCl₃CN (3.3 equiv), DBU (0.3 equiv), rt, 5 h, 86.3%.

Benzoylation of 1,2-*O*-ethylidene- β -D-mannopyranose⁷ (8) followed by hydrolysis with 90% CF₃COOH afforded the 3,4,6-tri-*O*-benzoyl-D-mannopyranose (10), subsequent acetylation with acetic anhydride in pyridine furnished the diacetate 11. Selective removal of the 1-*O*-acetyl group with benzylamine in THF (\rightarrow 12), and then treatment with CCl₃CN/DBU in dichloromethane afforded the glycosyl donor 13. (Scheme 2).

As shown in Scheme 3, allyl 6-*O*-acetyl-2,3,4-tri-*O*-benzoyl- α -D-mannopyranoside (14) was prepared by the Helferich reaction using 3 as the glycosyl donor and allyl alcohol as the acceptor.⁸ Selective removal of the acetyl group of 14 in



Scheme 3. Reagents and conditions: (a) Allyl alcohol (2 equiv.), TMSOTf (0.26 equiv), CH_2Cl_2 , rt, 3 h, 88%. (b) methanol/0.5% HCl, rt, 12–14 h, 93%–96%. (c) **5** (1.4 equiv), CH_2Cl_2 , TMSOTf (0.08 equiv), rt, 3 h, 86.5%. (d) **13** (1.4 equiv), CH_2Cl_2 , TMSOTf (0.16 equiv), rt, 3 h, 85.7%. (e) **7** (5 equiv), CH_2Cl_2 , TMSOTf (0.3 equiv), rt, 3 h, 80.4%. (f) i. PdCl_2, CH_3OH , rt, 4 h; ii. CH_3OH satd with dry NH_3 , rt, 72 h, 70.2% (two steps).



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methanol solution containing 0.5% HCl gave the glycosyl acceptor 15 in a high yield. The disaccharide 16 was prepared using the "inverse Schmidt" strategy.⁹ Thus the glycosyl acceptor 15 and the catalyst TMSOT were mixed first in dry CH_2Cl_2 , and after stirring for 15 min, the glycosyl donor 5 in dry dichloromethane was added dropwise within 30 min in order to give 16 in a high yield. Selective removal of the acetyl group of **16** gave the glycosyl acceptor **17**, "inverse Schmidt" coupling of which with 2-O-acetyl-3,4,6-tri-O-benzoyl- α -D-mannopyranosyl trichloroacetimidate (13) afforded the trisaccharide 18 in 85.7% yield. The 1 H NMR spectrum of **18** showed one acetyl signal (δ 2.14), one allyl signal, both signals characteristic of the structure of the trisaccharide 18. Selective removal of the 2-O-acetyl group of the trisaccharide 18 gave the glycosyl accepter 19. The fully protected tetrasaccharide 20 was smoothly obtained using the "inverse Schmidt" method, again by coupling with 2,3,4,6-tetra-O-benzoyl- α -D-mannopyranosyl trichloroacetimidate (7). Deprotection of **20** using PdCl₂ in CH₃OH,¹⁰ followed by treatment with NH₃ in CH₃OH, gave the title mannotetraose 1. The bioassay of 1 is in process.

EXPERIMENTAL

General Methods. Optical rotations were determined at 25°C with a Perkin-Elmer Model 241-Mc automatic polarimeter. Melting points were determined with a "Mel-Temp" apparatus. ¹H NMR and ¹³C NMR spectra were recorded with Bruker ARX 400 spectrometers for solutions in CDCl₃. Chemical shifts are given in parts per million (ppm) downfield from internal Me₄Si. Mass spectra were recorded with a JMS-D300S mass spectrometer using a direct sample introduction technique. Thin-layer chromatography (TLC) was performed on Silica Gel HF₂₅₄ with detection by charring with 30% (v/v) H₂SO₄ in MeOH or in some cases by UV detection. Column chromatography was conducted by elution of a column (16 × 240, 18 × 300, 35 × 400 mm) of silica gel (100–200 mesh) with EtOAc - petroleum ether (60–90°C) as the eluent. Solutions were concentrated at < 60°C under diminished pressure.

1,6-Di-*O***-acetyl-2,3,4-tri-***O***-benzoyl-** α **-D-mannopyranose (3).** A solution of mannose **2** (15 g, 83.3 mmol) and trityl chloride (28 g, 100.5 mmol) in pyridine (100 mL) was stirred at 50°C for 32 h, at the end of which time TLC (4:1 EtOAc-methanol) indicated that the reaction was complete. The reaction mixture was cooled to 0°C, and then benzoyl chloride (46.5 mL, 400.9 mmol) was added dropwise for 30 min to keep the reaction temperature under 40°C. After 24 h, water (300 mL) was added to the reaction mixture, and stirring was continued for 30 min. The aq solution was extracted with CH₂Cl₂ (3 × 100 mL), the extract was washed with HCl (1 N) and saturated aqueous sodium bicarbonate, dried (Na₂SO₄) and concentrated to dryness. The residue without separation was dissolved in CH₂Cl₂ (50 mL), Ac₂O (50 mL) and AcOH (30 mL), the solution cooled to 10°C in an ice bath, and H₂SO₄ (8.8 mL) added dropwise over 20 min. After the addition





was complete, the ice bath was removed and the reaction was continued for 20 h at ambient temperature. The reaction solution was poured into ice water (400 mL), stirring was continued for an additional 15 min, and the aqueous solution was extracted with chloroform (3×100 mL). The combined chloroform extracts were carefully washed with 10% aq NaHCO₃ (3×60 mL), dried over Na₂SO₄, and concentrated to a syrup which was subjected to column chromatography with 4:1 petroleum ether-EtOAc as the eluent. Compound **3** was obtained as a syrup (34.2 g, 71.3%); [α]_D + 9.5° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.11–7.28 (m, 15 H, 3 PhH), 6.36 (d, 1 H, J_{1,2} = 2.0 Hz, H-1), 6.01 (t, 1 H, J_{3,4} = J_{4,5} = 10.2 Hz, H-4), 5.88 (dd, 1 H, J_{2,3} = 3.2 Hz, J_{3,4} = 10.2 Hz, H-3), 5.70 (dd, 1 H, J_{1,2} = 2.0 Hz, J_{2,3} = 3.2 Hz, H-2), 4.34 (m, 2 H, H-5, 6a), 4.27 (dd, 1 H, J_{6a,6b} = 13.4 Hz, J_{5,6b} = 4.2 Hz, H-6b), 2.28, 2.08 (2 s, 6 H, 2COCH₃); ¹³C NMR (100 MHz, CDCl₃): 170.0, 167.7 (2 C, 2 COCH₃), 165.1, 165.0, 164.6 (3 C, 3 COPh), 133.3–127.8 (Ph), 90.2 (C-1), 70.2, 69.3, 68.8, 65.9 (4 C, C-2, 3, 4, 5), 62.0 (C-6), 20.4, 20.1 (2 C, 2 COCH₃).

Anal. Calcd for C₃₁H₂₈O₁₁: C, 64.58; H, 4.89. Found: C, 64.67; H, 4.94.

6-O-Acetyl-2,3,4-tri-O-benzoyl-α-D-mannopyranose (4). A solution of compound **3** (5 g, 8.67 mmol) and benzylamine (3 mL, 27.4 mmol) in anhydrous THF (30 mL) was stirred at room temperature for 24 h, at the end of which time TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solution was concentrated. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound **4** as a syrup (4.0 g, 86.2%); $[\alpha]_D$ + 16.8° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.11–7.27 (m, 15 H, 3 Ph*H*), 5.98 (m, 2 H, H-3, 4), 5.71 (dd, 1 H, J_{1,2} = 1.6Hz, J_{2,3} = 2.4 Hz, H-2), 5.53 (d, 1 H, J_{1,2} = 1.6 Hz, H-), 4.55 (m, 1 H, H-5), 4.33 (m, 2 H, H-6a, 6b), 2.09(s, 3 H,COCH₃); ¹³C NMR (100 MHz, CDCI₃) ; 170.6 (COCH₃), 166.1, 165.2, 165.2 (3 C, 3 *C*OPh), 133.1–127.9 (Ph), 91.7 (C-1), 70.6, 69.4, 68.0, 66.5 (4 C, C-2, 3, 4, 5), 62.4 (C-6), 20.3 (*C*OCH₃).

Anal. Calcd for C₂₉H₂₆O₁₀: C, 65.17; H, 4.90. Found: C, 65.42; H, 4.94.

6-O-Acetyl-2,3,4-tri-O-benzoyl-α-D-mannopyranosyl trichloroacetimidate (5). A mixture of **4** (4.0 g, 7.48 mmol), trichloroacetonitrile (2.1 mL, 20.9 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.25 mL, 1.67 mmol) in dry dichloromethane (25mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **5** (4.47 g, 88.1%) as crystals; mp 88–91°C; $[\alpha]_D$ + 20.3° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.87 (s, 1 H, OC(NH)CCI₃), 8.12–7.27 (m, 15 H, 3 PhH), 6.55 (d, 1 H, J_{1,2} = 2.0 Hz, H-1), 6.06 (t, 1 H, J_{3,4} = J_{4,5} = 10.0 Hz, H-4), 5.93 (dd, 1 H, J_{2,3} = 3.2 Hz, J_{3,4} = 10.0 Hz, H-3), 5.90 (dd, 1 H, J_{1,2} = 2.0 Hz, J_{2,3} = 3.2 Hz, H-2), 4.49 (m, 1 H, H-5), 4.34 (m, 2 H, H-6a, 6b), 2.07 (s, 3 H, COCH₃); ¹³C NMR (100 MHz, CDCI₃): 170.0 (*C*OCH₃), 165.0, 164.6, 159.4 (4 C, 3 *C*OPh, *C*(NH)CCI₃), 133.3–127.9 (Ph), 94.1 (C-1), 90.1 (*C*CI₃), 70.8, 69.2, 68.3, 65.7 (4 C, C-2, 3, 4, 5), 61.9 (C-6), 20.1 (*C*OCH₃).

Anal. Calcd for C₃₁H₂₆NO₁₀CI₃: C, 54.84; H, 3.86. Found: C, 54.72; H, 3.90.

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2,3,4,6-Tetra-*O***-benzoyl-** α **-D-mannopyranosyl trichloroacetimidate** (7). A mixture of 2,3,4,6-tetra-*O*-benzoyl- α -D-mannopyranose (**6**)¹¹ (3.5 g, 5.87 mmol), trichloroacetonitrile (1.9 mL, 18.9 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.20 mL, 1.34 mmol) in dry dichloromethane (30 mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give 7 (3.72 g, 85.6%) as crystals; mp 132–134°C; [α]_D +48.3° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.86 (s, 1 H, OC(N*H*)CCI₃), 8.10–7.26 (m, 20 H, 4 Ph*H*), 6.57 (d, 1 H, J_{1,2} = 1.3 Hz, H-1), 6.23 (t, 1 H, J_{3,4} = J_{4,5} = 10.0 Hz, H-4), 5.99–5.94 (, 2 H, H-2, 3), 4.72 (dd, 1 H, J_{5,6a} = 2.4 Hz, J_{6b,6a} = 12.3 Hz, H-6b), 4.62 (m, 1 H, H-5), 4.50 (dd, 1 H, J_{5,6b} = 4.0 Hz, J_{6b,6a} = 12.3 Hz, H-6b); ¹³C NMR (100 MHz, CDCI₃), 165.5, 165.0, 164.9, 164.6 (4 C, 4 COPh), 159.0 (*C*(NH)CCI₃), 133.2–127.9 (Ph), 94.2 (C-1), 71.1, 69.4, 68.4, 65.6 (4 C, C-2, 3, 4, 5), 61.95 (C-6).

Anal. Calcd for C₃₆H₂₈NO₁₀CI₃: C, 58.35; H, 3.81. Found: C, 58.44; H, 3.76.

3,4,6-Tri-*O*-benzoyl-**1,2**-*O*-(*S*-ethylidene)-β-D-mannopyranose (9). 1.2-O-(S-ethylidene)- β -D-mannopyranose (8)⁷ (6.4 g, 31.1 mmol) in pyridine (30 ml) was cooled to 0°C, and then benzoyl chloride (11.9 mL, 102.5mmol) was added dropwise for 30 min to keep the reaction temperature under 40°C. After 24 h, water (300 mL) was added to the reaction mixture, and stirring was continued for 30 min. The aq solution was extracted with CH_2Cl_2 (3 \times 100 mL), the extract was washed with HCl (1 N) and then saturated sodium bicarbonate, dried (Na₂SO₄) and concentrated to dryness. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **9** (15.3 g, 94.8%) as crystals; mp 123–125°C; $[\alpha]_D$ + 25.9° (c 1.0, CHCI₃); ¹H NMR (400MHz, CDCI₃): δ 8.04–7.25 (m, 15 H, 3 Ph*H*), 6.01 (t, 1 H, $J_{3,4} = J_{5,4} = 10.2$ Hz, H-4), 5.80 (dd, 1 H, $J_{3,2} = 3.6$ Hz, $J_{4,3}$ =10.2 Hz, H-3), 5.51 (dd, 1 H, J_{1.2} = 2.0 Hz, J_{3.2} = 3.6 Hz, H-2), 5.45 (q, 1 H, J = 5.0 Hz, CH₃CH), 5.36 (d, 1 H, J_{2.1} = 2.0 Hz, H-1), 4.70 (m, 1 H, H-5), 4.50–4.44 $(m, 2 H, H-6, 6'), 1.57 (d, 3 H, J = 5.0 Hz, CH_3CH); {}^{13}C NMR (100 MHz, CDCl_3),$ 165.7, 165.5, 164.8 (3 C, 3 COPh), 133.0–127.8 (Ph), 104.4 (CHCH₃), 96.3 (C-1), 77.06, 71.4, 71.0, 66.3 (4 C, C-2, 3, 4, 5), 62.9 (C-6), 21.1 (CHCH₃).

Anal. Calcd for C₂₉H₂₆O₉: C, 67.18; H, 5.05. Found: C, 67.33; H, 5.00.

3,4,6-Tri-*O***-benzoyl-D-mannopyranose (10)**. A solution of compound **9** (7.2 g, 13.9 mmol) in 90% F₃CCOOH (30 mL) was stirred at room temperature for 4 h, at the end of which time TLC (1:1 petroleum ether-EtOAc) indicated that the reaction was complete. The reaction mixture was diluted with toluene (50 mL) and then concentrated to dryness. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound 10 as a crystalline mixture of α and β forms in a ratio of 2:1 (6.1 g, 89.0%); mp 99–101°C; [α]_D + 40.8° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.04–7.27 (m, 15 H, 3 PhH), 6.00 (t, 0.66 H, J_{3,4} = J_{5,4} = 9.8 Hz, H-4 of α anomer), 5.92 (t, 0.34 H, J_{3,4} = J_{5,4} = 9.9 Hz, H-4 of β anomer), 5.76 (dd, 0.66 H, J_{3,2} = 3.2 Hz, J_{4,3} = 9.8 Hz, H-2 of α anomer), 5.44 (dd, 0.34 H, J_{3,2} = 3.1 Hz, J_{4,3} = 9.7 Hz, H-2 of β anomer), 5.43 (d, 0.66 H, J_{2,1} = 1.0 Hz, H-1 of α anomer), 5.1 (d, 0.34 H, J_{2,1} = 1.1 Hz, H-1 of β anomer),





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4.67–4.37 (m, 4 H, H-2, 5, 6, 6'); ¹³C NMR (100 MHz, CDCl₃): 166.1, 165.5, 165.3 (3 C, 3 COPh), 133.3–127.9 (Ph), 94.0, (C-1), 72.2, 69.2, 68.0, 66.5 (4 C, C-2, 3, 4, 5), 63.2 (C-6).

Anal. Calcd for C₂₇H₂₄O₉: C, 65.85; H, 4.91. Found: C, 66.02; H, 4.95.

1,2-Di-*O***-acetyl-3,4,6-tri-***O***-benzoyl-\alpha-D-mannopyranose** (**1**). Acetylation of **10** (3 g, 6.1 mmol) with acetic anhydride (8 mL) in pyridine (10 mL) at room temperature for 4 h gave compound **11** in a quantitative yield as crystals; mp 115–117°C; [α]_D + 35.9° (*c* 1.0, CHCI₃); ¹H NMR (400MHz, CDCI₃): δ 8.05–7.27 (m, 15 H, 3 Ph*H*), 6.24 (d, 1 H, J_{2,1} = 2.0 Hz, H-1), 6.01 (t, 1 H, J_{3,4} = J_{5,4} = 10 Hz, H-4), 5.80 (dd, 1 H, J_{3,2} = 3.6 Hz, J_{4,3} = 10.2 Hz, H-3), 5.51 (dd, 1 H, J_{1,2} = 2.0 Hz, J_{3,2} = 3.6 Hz, H-2), 4.63 (m, 1 H, H-5), 4.49–4.43 (m, 2 H, H-6,6'), 2.26, 2.18 (2 s, 6 H, 2 COCH₃); ¹³C NMR (75 MHz, CDCI₃): 169.5, 168.2 (2C, 2 COCH₃), 166.0, 165.6, 165.3 (3C, 3 COPh), 133.6–128.3 (Ph), 90.6 (C-1), 70.8, 69.5, 68.7, 66.4 (4 C, C-2, 3, 4, 5), 62.8 (C-6), 21.0, 20.7 (2 C, 2 COCH₃). Anal. Calcd for C₃₁H₂₈O₁₁: C, 64.58; H, 4.89. Found: C, 64.70; H, 4.93.

2-O-Acetyl-3,4,6-tri-O-benzoyl-α-D-mannopyranose (12). A solution of compound **11** (8 g, 13.8 mmol) and benzylamine (6 mL, 54.9 mmol) in anhydrous THF (50 mL) was stirred at room temperature for 24 h, at the end of which time TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solution was concentrated. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound **12** as crystals (6.56g, 88.4%); mp 89–91°C; $[\alpha]_D$ + 39.8° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.07–7.34 (m, 15 H, 3 Ph*H*), 5.96 (t, 1 H, J_{3,4} = J_{5,4} = 10 Hz, H-4), 5.87 (dd, 1 H, J_{3,2} = 3.2 Hz, J_{4,3} = 10.0 Hz, H-3), 5.50 (dd, 1 H, J_{1,2} = 2.0 Hz, J_{3,2} = 3.2 Hz, H-2), 5.37 (d, 1 H, J_{2,1} = 2.0 Hz, H-1), 4.66 (dd, 1 H, J_{6,6'} = 12.0 Hz, J_{5,6} = 2.8 Hz, H-6), 4.60 (m, 1 H, H-5), 4.44 (dd, 1 H, J_{6,6'} = 12.0 Hz, J_{5,6'} = 4.4 Hz, H-6), 2.18 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCl₃), 169.6 (COCH₃), 165.9, 165.1 (3 C, 3 COPh), 132.9–127.9 (Ph), 91.7 (C-1), 70.0, 69.1, 68.4, 66.6 (4 C, C-2, 3, 4, 5), 62.7 (C-6), 20.3 (COCH₃).

Anal. Calcd for C₂₉H₂₆O₁₀: C, 65.17; H, 4.90. Found: C, 65.02; H, 4.85.

2-O-Acetyl-3,4,6-tri-O-benzoyl-α-D-mannopyranosyl trichloroacetimidate (13). A mixture of **12** (4.9 g, 9.17 mmol), trichloroacetonitrile (3.0 mL, 30.0 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.40 mL, 2.7 mmol) in dry dichloromethane (40 mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **13** (5.37 g, 86.3%) as crystals; mp 87–90°C; $[\alpha]_D$ + 36.7° (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.82 (s, 1 H, OC(NH)CCI₃), 8.04–7.36 (m, 15 H, 3 Ph*H*), 6.42 (d, 1 H, J_{1,2} = 2.0 Hz, H-1), 6.03 (t, 1 H, J_{3,4} = J_{4,5} = 10.0 Hz, H-4), 5.85 (dd, 1 H, J_{2,3} = 3.4 Hz, J_{3,4} = 10.0 Hz, H-3), 5.71 (dd, 1 H, J_{1,2} = 2.0 Hz, J_{2,3} = 3.4 Hz, H-2), 4.64 (dd, 1 H, J_{6,6'} = 12.8 Hz, J_{5,6} = 7.0 Hz, H-6), 4.56 (m, 1 H, H-5), 4.46 (dd, 1 H, J_{6,6'} = 12.8 Hz, J_{5,6'} = 2.8 Hz, H-6'), 2.17 (s, 3H, COCH₃); ¹³C NMR (100 MHz, CDCI₃), 169.5 (COCH₃), 165.9, 165.1, 165.0,

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164.9(4 C, 3 COPh, C(NH)CCl₃), 133.2–127.9 (Ph), 95.7 (C-1), 91.7 (C(NH)CCl₃), 69.9, 69.1, 68.4, 66.6 (4 C, C-2, 3, 4, 5), 62.7 (C-6), 20.3 (COCH₃). Anal. Calcd for C₃₁H₂₆NO₁₀CI₃: C, 54.84; H, 3.86. Found: C, 54.69; H, 3.98.

Allyl 6-O-Acetyl-2,3,4-tri-O-benzoyl- α -D-mannopyranoside (14). A solution of 3 (2.5 g, 4.3 mmol) and allyl alcohol (0.6 mL, 8.8 mmol) in dry dichloromethane (40 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and then TMSOTf (0.2 mL, 1.1 mmol) was added dropwise. After 1 h the reaction mixture was diluted with dichloromethane (50 mL) and washed with a satd sodium hydrogen carbonate solution (15 mL). The organic layer was dried with Na₂SO₄ and concentrated *in vacuo*. Purification of the residue by flash chromatography (3:1 petroleum ether-EtOAc) gave 14 as a syrup (2.19 g, 88%); $[\alpha]_{D}$ + 15.9° (c 1.0, CHCI₃); ¹H NMR (400MHz, CDCI₃): δ 8.11–7.26 (m, 15 H, 3 PhH), 6.12–5.91 (m, 3 H, H-3, 4, $CH_2CH=CH_2$), 5.68 (dd, $J_{1,2} = 2.0$ Hz, $J_{3,2} = 2.8$ Hz, H-2), 5.40 (dd, 1 H, ²J = 1.6Hz, ³J_{trans} = 17.2 Hz, CH₂CH=CH₂), 5.30 (dd, 1 H, ${}^{2}J = 1.6$ Hz, ${}^{3}J_{cis} = 10.4$ Hz, CH₂CH=CH₂), 5.14 (d, J_{1,2} = 2.0 Hz, H-1), 4.47–4.27 (m, 4 H, H-6, 6', CH₂CH=CH₂), 4.17 (m, 1 H, H-5), 2.10 (s, 3 H,COCH₃); ¹³C NMR (100 MHz, CDCl₃), 170.1 (COCH₃), 165.7, 165.6, 165.5 (3 C,3 COPh), 133.7–128.4 (Ph, CH₂CH=CH₂), 118.7 (CH₂CH=CH₂), 96.7 (C-1), 70.5, 70.0, 69.0, 68.7, 66.9 (5 C, C-2, 3, 4, 5, CH₂CH=CH₂), 62.8 (C-6), 20.8 $(COCH_3).$

Anal. Calcd for C₃₂H₃₀O₁₀: C, 66.89; H, 5.26. Found: C, 66.93; H, 5.31.

Allyl 2,3,4-Tri-O-benzoyl- α -D-mannopyranoside (15). A solution of 14 (1.6 g, 2.8 mmol) in methanol solution (80 mL) containing 0.5 % HCl was stirred at room temperature for 12 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and CH_2Cl_2 , then the organic layer dried over Na_2SO_4 , and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 15 as a syrup (1.42 g, 96%); $[\alpha]_D$ + 14.2° (c 1.0, CHCI₃); ¹H NMR (400MHz, CDCI₃): δ 8.12–7.24 (m, 15 H,3 Ph*H*), 6.03–5.98 (m, 2 H, H-3, CH₂CH=CH₂), 5.85 (t, 1 H, $J_{3,4} = J_{4,5} = 10.0$ Hz, H-4), 5.69 (dd, $J_{1,2} = 1.6$ Hz, $J_{2,3} = 3.2$ Hz, H-2), 5.40 (dd, 1 H, ²J = 1.6 Hz, ³J_{trans} = 17.2 Hz, CH₂CH=CH₂), 5.29 (dd, 1 H, ${}^{2}J$ = 1.6Hz, ${}^{3}J_{cis}$ = 10.4 Hz, $CH_2CH=CH_2$), 5.16 (d, $J_{1,2} = 1.6$ Hz, H-1), 4.34–4.16 (m, 2 H, $CH_2CH=CH_2$), 4.11 (m, 1 H, H-5), 3.83–3.79 (m, 2 H, H-6,6'); ¹³C NMR (100 MHz, CDCl₃). 166.6, 165.6, 165.5 (3 C, 3 COPh), 133.7–128.3 (Ph, CH₂CH=CH₂), 118.5 (Ph, CH₂CH=CH₂), 96.8 (C-1), 71.0, 70.7, 69.6, 68.8, 67.3 (5 C, C-2, 3, 4, 5, $CH_2CH=CH_2$), 61.3 (C-6).

Anal. Calcd for C₃₀H₂₈O₉: C, 67.66; H, 5.30. Found: C, 67.83; H, 5.37.

Allyl 6-O-Acetyl-2,3,4-tri-O-benzoyl- α -D-mannopyranosyl- $(1 \rightarrow 6)$ -2,3,4tri-O-benzoyl- α -D-mannopyranoside (16). A solution of 15 (0.37 g, 0.69 mmol) and TMSOTf (10 µL, 0.055mmol) in dry dichloromethane (6 mL) was stirred with





dried molecular sieves (4 A, 0.4 g) under nitrogen for 15 min, and then 5 (0.66 g, 0.97 mmol) in dichloromethane (4 mL) was added dropwise for 20 min. After 3 h the reaction mixture was diluted with dichloromethane (30 mL) and washed with satd sodium hydrogen carbonate solution (5 mL). The organic layer was dried with Na₂SO₄ and concentrated in vacuo. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 16 as a syrup (0.63 g, 86.5%); $[\alpha]_D$ + 35.3° (c 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.08–7.27 (m, 30 H, 6 PhH), $6.07-5.91 \text{ (m, 5 H, H-3_A, 3_B, 4_A, 4_B, CH_2CH=CH_2)}, 5.77 \text{ (dd, 1 H, J}_{1,2} = 1.5 \text{ Hz},$ $J_{2,3} = 3.2 \text{ Hz}, \text{H-2}_{A}$), 5.74 (dd, 1 H, $J_{1,2} = 1.5 \text{ Hz}, J_{2,3} = 3.2 \text{ Hz}, \text{H-2}_{B}$), 5.50 (dd, 1 H, ${}^{2}J = 1.6$ Hz, ${}^{3}J_{trans} = 17.2$ Hz, CH₂CH=CH₂), 5.35 (dd, 1 H, ${}^{2}J = 1.6$ Hz, ${}^{3}J_{cis} =$ 10.4 Hz,), 5.18 (d, 1 H, $J_{1,2} = 1.5$ Hz, H-1_A), 5.13 (d, 1 H, $J_{1,2} = 1.5$ Hz, H-1_B), $4.45-3.76 (m, 8 H, H-5_A, 5_B, 6_A, 6_b, 6_A', 6_B', CH_2CH=CH_2), 1.92 (s, 3 H, COCH_3);$ ¹³C NMR (100 MHz, CDCl₃), 169.5 (COCH₃), 164.6, 164.5, 164.2, 164.1 (6 C, 6 COPh), 132.6–127.3 (Ph, CH₂CH=CH₂). 117.6 (CH₂CH=CH₂), 96.4, 95.7 (2 C, C-1_A, 1_B), 69.5, 69.1, 68.8, 68.5, 67.9, 67.7, 65.9, 65.7, 61.5 (11 C, C-2_A, 2_B, 3_A, 3_B, 4_A, 4_B, 5_A, 5_B, 6_A, 6_B, CH₂CH=CH₂), 19.5 (COCH₃).

Anal. Calcd for C₅₉H₅₂O₁₈: C, 67.55; H, 5.00. Found: C, 67.34; H, 4.96.

Allyl 2,3,4-Tri-O-benzoyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-ben $zoyl-\alpha$ -D-mannopyranoside (17). A solution of 16 (0.55 g, 0.52 mmol) in methanol solution (25 mL) containing 0.5 % HCl was stirred at room temperature for 14 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and CH_2Cl_2 , then the organic layer dried over Na_2SO_4 , and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 17 as a syrup (0.5 g, 95.6%); $[\alpha]_{D}$ + 31.0° (c 1.5, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.17–7.26 (m, 30 H, 6 PhH), 6.06–5.80 (m, 5 H, H-3_A, 3_B, 4_A, 4_B, CH₂CH=CH₂), 5.76–5.73 (m, 2 H, H-2_A, 2_B), 5.34 (dd, 1 H, ${}^{2}J = 1.6$ Hz, ${}^{3}J_{trans} = 17.2$ Hz, CH₂CH=CH₂), 5.50 (dd, 1 H, ${}^{2}J = 1.6$ Hz, ${}^{3}J_{cis}$ = 10.4 Hz,), 5.17 (d, 1 H, $J_{1,2}$ = 1.6 Hz, H-1_A), 5.14 (d, 1 H, $J_{1,2}$ = 1.6 Hz, H- $1_{\rm B}$), 4.44–3.55 (m, 8 H, H- $5_{\rm A}$, $5_{\rm B}$, $6_{\rm A}$, $6_{\rm B}$, $6_{\rm A}'$, $6_{\rm B}'$, $CH_2CH=CH_2$); ¹³C NMR (100 MHz, CDCl₃), 166.7, 165.7, 165.5, 165.4, 165.2 (6 C, 6 COPh), 133.7–128.3 (Ph, $CH_2CH=CH_2$), 118.6 ($CH_2CH=CH_2$), 97.8, 96.7 (2 C, C-1_A, 1_B), 70.9, 70.6, 70.4, 70.2, 69.5, 68.9, 67.1, 67.0, 66.7, 61.0 (11 C, C-2_A, 2_B, 3_A, 3_B, 4_A, 4_B, 5_A, 5_B, 6_A, $6_{\rm B}, CH_2CH=CH_2$).

Anal. Calcd for C₅₇H₅₀O₁₇: C, 67.99; H, 5.00. Found: C, 68.11; H, 5.02.

Allyl 2-*O*-Acetyl-3,4,6-tri-*O*-benzoyl- α -D-mannopyranosy- $(1\rightarrow 6)$ -2,3,4-tri-*O*-benzoyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-*O*-benzoyl- α -D-mannopyranoside (18). A solution of 17 (0.25 g, 0.25 mmol) and trimethylsilyl trifluoromethanesulfonate (8 μ L, 0.04 mmol) in dry dichloromethane (20 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and 13 (0.24 g, 0.35 mmol) in dry dichloromethane (10 mL) was added dropwise for 30 min. After 3 h the reaction mixture was diluted with dichloromethane (20 mL) and

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washed with satd sodium hydrogen carbonate solution (8 mL). The organic layer was dried with Na₂SO₄ and concentrated in vacuo. Purification of the residue by column chromatography (2:1 petroleum ether-EtOAc) gave **18** as a syrup (0.32 g, 85.7%); $[\alpha]_D + 28.7^{\circ}$ (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.10–7.25 (m, 45 H, 9 PhH), 6.21–5.88 (m, 7 H, H-3_A, 3_B, 3_C, 4_A, 4_B, 4_C, CH₂CH=CH₂), 5.84 (dd, 1 H, J_{1,2} = 1.6 Hz, J_{2,3} = 3.2 Hz, H-2_A), 5.79 (dd, 1 H, J_{1,2} = 1.6 Hz, J_{2,3} = 3.2 Hz, H-2_A), 5.79 (dd, 1 H, J_{1,2} = 1.6 Hz, J_{2,3} = 3.2 Hz, H-2_B), 5.44 (dd, 1 H, ²J = 1.6 Hz, ³J_{trans} = 17.2 Hz, CH₂CH=CH₂), 5.37 (dd, 1 H, J_{1,2} = 1.6 Hz, J_{2,3} = 3.2 Hz, H-2_C), 5.26 (dd, 1 H, ²J = 1.6 Hz, ³J_{cis} = 10.4 Hz, CH₂CH=CH₂), 5.20 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_A), 5.16 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_B), 4.73 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_C), 4.43–4.37 (m, 11 H, H-5_A, 5_B, 5_C, 6_A, 6_B, 6_C, 6_A', 6_B', 6_C', CH₂CH=CH₂), 2.07 (s, 3 H, COCH₃); ¹³C NMR (100 MHz, CDCl₃), 169.5 (COCH₃), 166.0, 165.7, 165.5, 165.3, 165.2 (9 C, 9 COPh), 133.4–128.3 (Ph, CH₂CH=CH₂), 118.8 (CH₂CH=CH₂), 98.0, 97.2, 96.9 (3 C, C-1_A, 1_B, 1_C), 20.7 (COCH₃).

Anal. Calcd for C₈₆H₇₄O₂₆: C, 67.80; H, 4.90. Found: C, 68.04; H, 4.96.

Allyl 3,4,6-Tri-O-benzoyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- α -D-mannopyranosyl- $(1 \rightarrow 6)$ -2,3,4-tri-O-benzoyl- α -D-mannopyranoside (19). A solution of 18 (0.37 g, 0.24 mmol) in methanol solution (25 mL) containing 0.5 % HCl was stirred at room temperature for 14 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and CH₂Cl₂, then the organic layer dried over Na₂SO₄, and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 19 as a syrup (0.34 g, 93.5%); $[\alpha]_{\rm D}$ + 23.0° (c 1.5, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.09–7.26 (m, 45 H, 9 PhH), 6.19–5.70 (m, 10 H, H-2_A, 2_B, 2_C, 3_A, 3_B, 3_C, 4_A, 4_B, $4_{\rm C}$, CH₂CH=CH₂), 5.48 (dd, 1 H, ²J = 1.6 Hz, ³J_{trans} = 17.2 Hz, CH₂CH=CH₂), $5.29 (dd, 1 H, {}^{2}J = 1.6 Hz, {}^{3}J_{cis} = 10.4 Hz, CH_{2}CH = CH_{2}), 5.20 (d, 1 H, J_{1.2} = 1.6 Hz)$ Hz, H-1_A), 5.17 (d, 1 H, $J_{1,2} = 1.6$ Hz, H-1_B), 4.75 (d, 1 H, $J_{1,2} = 1.6$ Hz, H-1_C), 4.43–4.37 (m, 11 H, H-5_A, 5_B, 5_C, 6_A , 6_B , 6_C , $6_A'$, $6_B'$, $6_C'$, $CH_2CH=CH_2$); ¹³C NMR (100 MHz, CDCl₃), 166.1, 165.8, 165.7, 165.6, 165.5, 165.4, 165.3, 165.2, 165.1 (9 C, 9 COPh), 133.6–27.7 (Ph, CH₂CH=CH₂), 118.5 (CH₂CH=CH₂), 97.7, 96.8, 96.4 (3 C, C-1_A, 1_B, 1_C).

Anal. Calcd for C₈₄H₇₂O₂₅: C, 68.10; H, 4.90. Found: C, 67.99; H, 4.94.

Allyl 2,3,4,6-Tetra-O-benzoyl- α -D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-Obenzoyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- α -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- α -D-mannopyranoside (20). A solution of 19 (0.21 g, 0.14 mmol) and trimethylsilyl trifluoromethanesulfonate (8 μ L, 0.04 mmol) in dry dichloromethane (20 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and then 7 (0.52 g, 0.70 mmol) in dry dichloromethane (10 mL) was added dropwise for 30 min. After 3 h the reaction mixture was diluted with dichloromethane (20 mL) and washed with satd sodium hydrogen carbonate solution (8 mL). The organic layer was dried over Na₂SO₄ and





concentrated *in vacuo*. Purification of the residue by column chromatography (2:1 petroleum ether-EtOAc) gave **20** as a syrup (0.23 g, 80.4%); $[\alpha]_D + 14.7^{\circ}$ (*c* 1.0, CHCI₃); ¹H NMR (400 MHz, CDCI₃): δ 8.04–7.18 (m, 65 H, 13 Ph*H*), 6.21–5.80 (m, 13 H, H-2_A, 2_B, 2_C, 2_D, 3_A, 3_B, 3_C, 3_D, 4_A, 4_B, 4_C, 4_D, CH₂CH=CH₂), 5.45 (dd, 1 H, ²J = 1.6 Hz, ³J_{trans} = 17.2 Hz, CH₂CH=CH₂), 5.28 (dd, 1 H, ²J = 1.6 Hz, ³J_{cis} = 10.4 Hz, CH₂CH=CH₂), 5.24 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_A), 5.22 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_B), 5.05 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_C), 4.87 (d, 1 H, J_{1,2} = 1.6 Hz, H-1_D), 4.43–4.37 (m, 14 H, H-5_A, 5_B, 5_C, 5_D, 6_A, 6_B, 6_C, 6_D, 6_A', 6_B', 6_C', 6_D', CH₂CH=CH₂); ¹³C NMR (100 MHz, CDCl₃), 166.2, 165.9, 165.7, 165.6, 165.5, 165.4, 165.2, 165.0, 164.8 (13 C, 13 COPh), 133.4–128.3 (Ph, CH₂CH=CH₂), 118.4 (CH₂CH=CH₂), 99.7, 98.1, 98.0, 97.6 (4 C, C-1_A, 1_B, 1_C, 1_D).

Anal. Calcd for C₁₁₈H₉₈O₃₄: C, 68.80; H, 4.79. Found: C, 68.97; H, 4.74.

α-D-Mannopyranosyl-(1→2)-α-D-mannopyranosyl-(1→6)-α-Dmannopyranosyl-(1→6)-D-mannopyranose (1). A mixture of compound 20 (0.1 g, 0.048 mmol) and PdCl₂ (0.02 mg) in dry methanol (10 mL) was stirred vigorously for 4 h at room temperature, TLC (1:1 petroleum ether-EtOAc) indicated that the reaction was complete, then filtered through Celite. The filtrate was concentrated to dryness. The resulting compound was dissolved in methanol (20 mL) satd with dry NH₃ and stirred at room temperature for 72 h, at the end of which time TLC (1:1.5 Methanol-EtOAc) indicated that the reaction was complete. The solution was concentrated to dryness and was chromatographed (methanol) on a column of Sephadex LH-20 to afford 1 (0.023 g, 70.2%) as an amorphous mass; [α]_D + 3.1° (*c* 0.1, MeOH); ¹H NMR (400 MHz, D₂O): δ 5.09, 5.01, 4.91, 4.89 (4 d, 4 H, J_{1,2} = 1.5 Hz, H-1_A, 1_B, 1_C, 1_D); ¹³C NMR (100 MHz, D₂O), δ 100.3, 100.1, 99.8, 98.6 (4 C, C-1_A, 1_B, 1_C, 1_D). ESMS Calcd for C₂₄H₄₂O₂₁: 666.58 (M). Found: 665.4 (M-H).

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REFERENCES

- Gorin, P.A.J.; Spencer, J.F.T. Structural Chemistry of Fungal Polysaccharides. Adv. Appl. Microbiol. 1970, 13, 25–42.
- Bovina, E.V.; Deryabin, V.V.; Gagloev, V.N.; Serebryakov, N.G. Carbon-13 NMR Study of Mannans Produced by *Candida maltosa* and *Candida tropicalis*. Prikl. Biokhim. Mikrobiol. **1988**, 24 (2), 218–225.
- Suzuki, M.; Fukazawa, Y. Immunochemical Characterization of *Candida albicans* Cell Wall Antigens: Specific Determinant of *Candida albicans* Serotype A Mannan. Microbiol. Immunol. **1982**, *26* (5), 387–402.

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- 4. Kogan, G.; Pavliak, V.; Masler, L. Structural Studies of Mannans from the Cell Wall of the Pathogenic Yeasts *Candida albicans* Serotypes A and B and *Candida parapsilosis*. Carbohydr. Res. **1988**, *172* (2), 243–253.
- 5. Kogan, G.; Pavliak, V.; Sandula, J.; Masler, L. Novel Structure of the Cellular Mannan of the Pathogenic Yeast *Candida Krasei*. Carbohydr. Res. **1988**, *184*, 171–182.
- Jiménez-Barbero, J.; Prieto, A.; Gómez-Miranda, B.; Leal, J.A.; Bernabé, M. Chemical Structure of Fungal Cell-Wall Polysaccharides Isolated from *Microsporum gypseum* and Related Species of *Microsporum* and *Trychophyton*. Carbohydr. Res. 1995, 272 (1), 121–128.
- Betaneli, V.I.; Ovchinnikov, M.V.; Backinowsky, L.V.; Kochetkov, N.K. A Convenient Synthesis of 1,2-*O*-Benzylidene and 1,2-*O*-Ethylidene Derivatives of Carbohydrates. Carbohydr. Res. 1982, *107* (2), 285–291.
- 8. Helferich, B.; Schmitz-Hillebrecht, E. A New Method for the Synthesis of Glucosides of the Phenols. Ber. **1933**, *66B*, 378–383.
- 9. Schmidt, R.R.; Toepfer, A. Glycosylation with Highly Reactive Glycosyl Donors: Efficiency of the Inverse Procedure. Tetrahedron Lett. **1991**, *32* (28), 3353–3356.
- a) Boss, R.; Scheffold, R. Cleavage of Allyl Ether with Palladium/Carbon. Angew. Chem. 1976, 88 (17), 578–579; b) Ogawa, T.; Nakabayashi, S. Synthetic Studies on Cell-Surface Glycans. Part 8. Synthesis of a Hexasaccharide Unit of a Complex Type of Glycan Chain of a Glycoprotein. Carbohydr. Res.1981, 93 (1), C1–C5.
- 11. Nikolaev, A.V.; Ivanova, I.A. Shibaev, V.N.; Kochetkov, N.K. Application of the Hydrogenphosphonate Approach in the Synthesis of Glycosyl Phosphosugar Linked through Secondary Hydroxyl Groups. Carbohydr. Res. **1990**, *204*, 65–78.

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